Endocrine Research

Thyroglobulin Is a Sensitive Measure of Both Deficient and Excess Iodine Intakes in Children and Indicates No Adverse Effects on Thyroid Function in the UIC Range of 100–299 μ g/L: A UNICEF/ICCIDD Study Group Report

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Context: The median urinary iodine concentration (UIC) is a biomarker of iodine intake. According to the World Health Organization, a median UIC in the range $100-199~\mu g/L$ indicates adequate and $200-299~\mu g/L$ more than adequate intake. Thyroglobulin (Tg) may be a promising functional biomarker of both iodine deficiency and excess.

Objectives: Using a standardized dried blood spots-Tg assay in children, we evaluated the Tg response to both low- and high-iodine intake and estimated the population cutoff point for iodine deficiency or excess. Also, we compared thyroid functions within the UIC ranges of 100-199 vs $200-299 \mu g/L$.

Design and Setting: We conducted a cross-sectional study in primary schools in 12 countries.

Subjects: Subjects were 6 to 12 years old (n = 2512).

Main Outcome Measures: We measured UIC, TSH, total T₄, Tg, and thyroid antibodies.

Results: Over a range of iodine intakes from severely deficient to excessive, Tg concentrations showed a clear U-shaped curve. Compared with iodine-sufficient children, there was a significantly higher prevalence of elevated Tg values in children with iodine deficiency (UIC <100 μ g/L) and iodine excess (UIC >300 μ g/L). There was no significant change in the prevalence of elevated Tg, TSH, T₄, or thyroid antibodies comparing children within the UIC ranges of 100–199 vs 200–299 μ g/L.

Conclusions: In school-aged children, 1) Tg is a sensitive indicator of both low and excess iodine intake; 2) a median Tg of <13 μ g/L and/or <3% of Tg values >40 μ g/L indicates iodine sufficiency in the population; 3) the acceptable range of median UIC in monitoring iodized salt programs could be widened to a single category of sufficient iodine intake from 100 to 299 μ g/L. (*J Clin Endocrinol Metab* 98: 0000–0000, 2013)

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Abbreviations: Ab, antibody; CI, confidence interval; DBS, dried whole-blood spots; Tg, thyroglobulin; TPO, thyroperoxidase; UIC, urinary iodine concentration; USI, universal salt iodization

There has been significant global progress against iodine deficiency due to the introduction of universal salt iodization (USI) in deficient areas (1). USI programs require careful monitoring because both iodine deficiency and iodine excess have adverse health effects (2). The two indicators commonly used to monitor iodine nutrition are measures of exposure: household coverage with iodized salt and the median urinary iodine concentration (UIC) (3). Neither of these assesses changes in thyroid function in response to varying iodine intake. Yet the ultimate goal of USI is to correct thyroid dysfunction caused by iodine deficiency to ensure optimal health. Thus, a functional biomarker of thyroid status, responsive to both low and high intakes of iodine, would improve USI monitoring.

Thyroglobulin (Tg) is synthesized only in the thyroid and is the most abundant intrathyroidal protein (4, 5). Transcytosis of Tg-containing endosomes across the thyrocyte results in release of small amounts of Tg into the circulation (6, 7). Serum Tg is elevated in iodine-deficient areas due to TSH hyperstimulation and thyroid hyperplasia (8). In intervention studies in adults, serum Tg is a more sensitive indicator than TSH or T_4 in measuring response to iodized oil (9–11), potassium iodide (12), and iodized salt (13). In intervention studies in children, Tg falls rapidly with iodine repletion (14).

Commercially available assays measure serum Tg, which requires venipuncture, centrifugation, and frozen sample transport, which is difficult in remote areas. We therefore developed an assay for Tg for dried whole-blood spots (DBS), simplifying collection and transport (15, 16), and established an international reference range (4–40 µg/L) in iodine-sufficient 5- to 14-year-old children (16). DBS Tg is a sensitive biomarker of improved thyroid function after iodine repletion (15, 16). World Health Organization (WHO)/UNICEF/International Council for the Control of Iodine Deficiency Disorders (ICCIDD) now recommend DBS Tg for the monitoring of iodine status in school-aged children (3).

However, several issues should be resolved before serum or DBS Tg can be widely adopted as a functional biomarker for monitoring iodine status. It is unclear whether Tg increases with increasing severity of iodine deficiency in children or whether Tg increases with excess iodine intake. It is also unknown how frequently anti-Tg antibodies (Ab) occur in children at varying iodine intakes; if common, these could confound measures of Tg. Finally, for population iodine monitoring, the cutoff point for median Tg that identifies iodine deficiency (or excess) remains uncertain. Therefore, our study aims were to, in school-age children, 1) evaluate the response of the standardized DBS-Tg assay to low and high intakes of iodine, 2) estimate the population cutoff point for using DBS-Tg

to define iodine deficiency, and 3) assess thyroid function over the range of iodine intakes currently defined by WHO/UNICEF/ICCIDD as adequate (median UIC, 100–199 μ g/L) and more than adequate (median UIC, 200–299 μ g/L) from iodized salt.

Subjects and Methods

Subjects

The international sample for this study included apparently healthy 6- to 12-year-old primary school children living in 12 countries: 2 in South America (Peru and Paraguay), 2 in Central Europe (Switzerland and Croatia), 2 in North Africa and the Eastern Mediterranean (Morocco and Bahrain), 2 in Sub-Saharan Africa (Tanzania and South Africa), 2 in Asia (Tajikstan and China), and 2 in Southeast Asia (Indonesia and the Philippines). These countries were selected to provide varying regional and ethnic representation. The study sites were selected to obtain varying iodine status and are not nationally representative; the data presented here should not be used for program purposes. In each country, 1 to 5 schools were selected to participate. The selection of the schools was purposeful; they were chosen 1) to represent specific areas of the country known to have iodine status that was deficient, sufficient, or more than adequate/excessive (based on WHO criteria for the median UIC) and 2) where there had been no recent change in iodine intake, that is, to represent customary, long-term iodine intakes. In nearly all areas, with the exception of one site in Indonesia with high iodine content in drinking water, the sources of dietary iodine were the local foods and variable amounts of iodized salt.

Recruitment was from primary schools at the middle to lower socioeconomic level. Exclusion criteria were 1) age <6 or >12 years, 2) chronic diseases, 3) use of chronic medications or iodine supplements, and 4) in females, pregnancy. With the relative precision for the 97th percentile for DBS-Tg specified at 3%-5% of the total length of the 95% reference range, and the estimated SD of DBS-Tg taken as 2.1 μ g/L (based on unpublished data from healthy Swiss children), we estimated a sample size of \sim 500 children would be required to obtain the required precision level in children with sufficient iodine intake (17). Due to the greater uncertainty on the variability of DBS-Tg in children with low and high iodine intake, we aimed to enroll a total sample size of roughly 2500 children, distributed evenly over the range of intake from deficient to excessive. Ethical committees approved the protocol at the Swiss Federal Institute of Technology, Zürich, Switzerland, and at each local institution involved in the study. Informed written consent was obtained from the parents and oral assent from the participating children. Data collection was carried out between 2006 and 2012.

Study design

The study design was cross-sectional. At the schools, height and weight were measured using standard anthropometric technique (18). For the measurements, children removed their shoes, emptied their pockets, and wore light indoor clothing. Heights were recorded to the nearest millimeter and weights to the nearest 100 g. Pubertal staging was not done. Spot urine samples were obtained from the children, aliquoted, and stored at -20° C until analysis. Whole blood from a finger stick was spotted onto filter

Table 1. Subject Characteristics, UIC, and DBS Tg, by Country

		Gender					
	n	(M/F)	Age, y ^a	Height, m ^a	Weight, kg ^a	UIC, μ g/L $^{ m b}$	Tg, μ g/L c
Morocco	248	120/128	9.2 ± 1.78	1.27 ± 0.13	25.9 ± 5.9	16 (1–95)	25.5 ± 44.2
Tajikistan	593	305/288	9.1 ± 0.73	1.26 ± 0.06	26.0 ± 3.3	52 (3–278)	10.9 ± 38.0
Switzerland	72	39/33	9.8 ± 1.40	1.40 ± 0.09	34.2 ± 7.0	137 (6-390)	10.5 ± 7.0
Philippines	230	120/110	10.1 ± 0.66	1.19 ± 0.10	24.9 ± 5.0	154 (8-706)	13.1 ± 12.8
Bahrain	147	74/73	9.6 ± 1.73	1.37 ± 0.11	33.5 ± 12.4	178 (43-701)	18.0 ± 13.6
Peru	125	78/47	9.4 ± 1.75	1.33 ± 0.11	32.5 ± 9.2	197 (22-890)	11.5 ± 12.6
Croatia	157	76/81	9.1 ± 1.44	1.40 ± 0.10	35.5 ± 10.5	205 (1–505)	11.3 ± 10.5
China	244	122/122	9.0 ± 2.03	1.37 ± 0.13	32.6 ± 11.5	235 (31-672)	12.6 ± 10.0
Indonesia	327	164/163	9.0 ± 0.83	1.27 ± 0.07	25.5 ± 6.1	235 (20-394)	9.8 ± 7.8
Paraguay	76	38/38	8.8 ± 1.78	1.33 ± 0.12	30.1 ± 8.5	257 (74-548)	13.6 ± 12.3
South Africa	120	61/59	9.3 ± 1.91	1.34 ± 0.13	33.3 ± 12.1	282 (38-758)	18.6 ± 26.1
Tanzania	173	95/78	8.7 ± 0.15	1.28 ± 0.11	25.3 ± 4.6	338 (6-1883)	17.6 ± 14.3
All	2512	1363/1251	9.4 ± 1.58	1.31 ± 0.25	29.0 ± 9.0	151 (1–1883)	13.3 ± 26.7

Abbreviations: F, female; M, male.

paper (grade 903; Schleicher & Schuell, Dassel, Germany) and allowed to dry at room temperature for 24 hours. They were then stored at 4°C in sealed low-density polyethylene bags until shipment to Zürich for analysis.

Laboratory analysis

UIC was measured using the Pino modification of the Sandell-Kolthoff reaction (19). External controls were provided by the EQUIP program (U.S. Centers for Disease Control and Prevention, Atlanta, Georgia). DBS were analyzed for TSH (DELFIA NeoTSH) (20) and total T_4 (Delfia Neonatal T_4 kit), both from PerkinElmer Life Sciences (Turku, Finland). Normal reference values are as follows: on whole blood, TSH of 0.2–3.7 mU/L, and on serum, total T_4 of 65–165 nmol/L. Tg-Ab and thyroperoxidase (TPO)-Ab were measured by RIA (RSR, Cardiff, United Kingdom) adapted in our laboratory for measurement on DBS (16). For the Tg-Ab assay, between- and within-assay coefficient of variation is 10.1% and 2.5%; for the TPO-Ab assay, between- and within-assay coefficient of variation is 12.4% and 2.1% (n = 145). Elevated Tg-Ab status was classified as greater than 10 U/ml; elevated TPO-Ab status was classified as greater than 12 II/m

For analysis of DBS-Tg, a two-site DELFIA serum Tg assay (PerkinElmer), adapted for DBS, was used (15). An advantage of two-site Tg assays is their lower cross-reactivity and improved specificity compared with one-site assays (4). The lyophilized Tg reference preparation of the Community Bureau of Reference of the Commission of the European Communities (CRM-457) was used to prepare calibrators for the DBS-Tg assay as described previously (15, 16).

Statistical analyses

Data processing and statistics were done using IBM SPSS statistics version 20. Only subjects with data on both UIC and thyroid function markers (Tg, TSH, and T_4) were included in the analysis. Non–normally distributed data were log-transformed for further analysis. For parameters including values between 0 and 1 (TSH and Tg), a constant of 1 was added to the values before transformation. Arithmetic mean \pm SD was used to report

normally distributed data, geometric mean \pm SD for data that were normally distributed after log-transformation, and median for data that were not normally distributed after log-transformation. Oneway ANOVA with post hoc Bonferroni correction was used to test differences between groups. Spearman correlations were calculated between UIC and thyroid function markers and the Loess smoothed line calculation (with 60% of points to fit) was used to describe the best fit of the thyroid function markers plotted against UIC. Significance was set at P < .05.

Results

General subject characteristics by country are shown in Table 1. Urine samples for the determination of UIC as well as DBS for the measurement of TSH, T₄, and Tg were available from 2512 children. Tg-Ab and TPO-Ab were measured on a subgroup of children (956 and 884, respectively). The median UIC of the entire sample was 151 μg/L; by country, Moroccan children had the lowest median UIC, 16 µg/L, whereas Tanzanian children had the highest, 338 μ g/L. The DBS Tg values by country are shown in Table 1; the overall geometric mean Tg concentration was $13.3 \pm 26.7 \,\mu\text{g/L}$, with the lowest concentration in Indonesia, $9.8 \pm 7.8 \mu g/L$, and the highest in Morocco, $25.5 \pm 44.2 \,\mu\text{g/L}$. The geometric mean TSH of all countries was 0.86 ± 0.81 mU/L, ranging from $0.61 \pm$ 0.21 mU/L in Switzerland to 1.45 \pm 0.86 mU/L in China. The arithmetic mean T_4 was 91.6 \pm 29.4 nmol/L, ranging from 62.2 ± 16.0 nmol/L in Paraguay to 114 ± 20.4 nmol/L in China.

Table 2 shows thyroid functions according to the WHO/UNICEF/ICCIDD categories of UIC used to classify iodine intake in a population of school-aged children. The distribution of UIC was well balanced: 958 children

^a Arithmetric mean \pm SD.

^b Median (minimum – maximum).

 $^{^{\}rm c}$ Geometric mean \pm SD.

Table 2. Thyroid Function in 6- to 12-Year-Old Children by WHO Categories of UIC

	UIC, μg/l	UIC, μg/L		
	<50	50-99.9		
lodine intake	Moderate to severe iodine deficiency	Mild iodine deficiency		
n	600	358		
TSH, mU/L	$0.87 \pm 1.27^{a,b}$	$0.78 \pm 0.43^{\circ}$		
T ₄ , nmol/L	$93.0 \pm 29.5^{a,b}$	97.4 ± 32.6^{a}		
Tg, μg/L	21.0 ± 44.1^{a}	$9.3 \pm 18.5^{b,c}$		
Tg-Ab, U/mL	$0.20 (0.01-1.57) (n = 240)^a$	$0.10 (0.10-16.9) (n = 91)^b$		
TPO-Ab, U/mL	$3.57 \pm 1.84 (n = 236)^a$	$4.17 \pm 2.35 (n = 76)^{a,b}$		

Results are shown as geometric mean \pm SD for TSH, Tg, and TPO-Ab; as arithmetric mean \pm SD for T₄; and as median (min-max) for Tg-Ab.

had a UIC in the deficient range, whereas 945 had a UIC in the range indicating more than adequate or excess iodine intake; 609 had a UIC in the range indicating adequate iodine intake. Significant group differences were found for TSH, T₄, and Tg, with the differences in Tg being most pronounced: Tg concentrations were highest in moderate to severe deficiency and in iodine excess (P < .05). Table 3 shows the prevalence of thyroid dysfunction in the sample by the UIC categories of iodine intake. There was a nonsignificant increase in subclinical and overt hypothyroidism at UIC <50 μ g/L (moderate to severe deficiency). The frequency of elevated Tg values was significantly higher in both iodine deficiency (UIC $<100 \mu g/L$) and in iodine excess (UIC > 300 μ g/L) (P < .05). Thyroid autoimmunity was extremely rare at all levels of iodine intake. To further examine the effect of high iodine intake on Tg, children in the iodine excess range were further divided in two groups: UIC 300-399 μ g/L (n = 248) and UIC >400 μ g/L (n = 188). The corresponding geometric mean Tg concentrations were 15.5 and 20.0 μg/L, respectively, and the percentages of elevated Tg ($>40 \mu g/L$) were 5.3% and 12.6%, respectively. For both groups, the values were significantly higher as compared with those from children in the UIC ranges of 100–199 and 200–299 µg/L (P < .01).

As expected, UIC was significantly positively correlated with TSH (r = 0.053, P = .008), and negatively with T_4 (r = -0.049, P = .015) and T_8 (r = -0.100, P < .001). However, the correlations were weak, and linear regression did not describe the data well, so Loess smoothed-line calculations were used for detailed analysis. Figures 1A, 2A, and 3A show the plots of T_4 and log TSH and T_8 against log UIC including the Loess smoothed line depicting the best fit. Because UIC is best applied as a population indicator, and to illustrate the influence of habitual iodine intake on thyroid function, bubble plots (Figures 1B, 2B, and 3B) were drawn of the median UIC for the 34 different school clusters plotted against the mean T_4 and the geo-

metric mean TSH and Tg of the clusters. The mean sample size of the schools was 74.

To examine the relationship between Tg and UIC over the middle range of adequate and more than adequate intakes, correlations were done between the median UIC values of the 19 school clusters with median UIC >100 and <300 μ g/L, and the corresponding mean Tg values for the clusters. There was no significant change in mean Tg within this UIC range (r = 0.327, P = .172). In the children from schools with median UIC >100 and <300 μ g/L (n = 1443), the geometric mean Tg concentration was 12.6 μ g/L (95% confidence interval [CI], 12.1–13.1) (median, 13.0; 95% CI, 12.6–13.4), and the 3rd and 97th percentiles (95% CI) were 2.9 (2.7–3.2) and 44.4 (39.3–47.7).

Discussion

The major finding of this study is that, over a range of iodine intake from severely deficient to excessive, Tg concentrations show a clear U-shaped curve (Figure 3, A and B). Compared with children with UIC in the adequate and more than adequate range (100–299 μ g/L), there was a higher prevalence of elevated Tg values in children with iodine excess (>300 μ g/L) and iodine deficiency (<100 μ g/L) (Table 3), and mean Tg values were significantly higher in children with UIC indicating moderate to severe deficiency and iodine excess (Table 2). These data suggest Tg could be used as a sensitive indicator in children not only of low iodine intake but also of excessive intake.

The DBS-Tg reference interval for iodine-sufficient, Tg-Ab-negative, euthyroid school-age children, using CRM-457 standardization, is $4-40~\mu g/L$ (16), nearly the same as the adult reference range for serum Tg when CRM-457 standardization is used, ie, approximately $3-40~\mu g/L$ (20). Therefore, if the percentage of children in a population with Tg concentrations above the upper reference value of $40~\mu g/L$ is greater than 3%, this suggests iodine deficiency

 $^{^{}a-c}$ Means not sharing a common superscript letter are significantly different from each other (one-way ANOVA with post hoc Bonferroni test; P < .05 was considered significant).

Table 2. Continued

UIC, μg/L							
200-299.9	>300						
More than adequate iodine intake 468	Excess iodine intake 477						
0.87 ± 0.59 ^{a,b} 89.4 ± 29.0 ^b	0.91 ± 0.70 ^b 88.7 ± 26.7 ^b						
$0.10 (0.10-109.2) (n = 170)^{b}$	17.4 ± 18.0^{d} $0.10 (0.10-69.4) (n = 198)^{b}$ $4.60 \pm 2.07 (n = 196)^{b}$						
	200–299.9 More than adequate iodine intake 468 $0.87 \pm 0.59^{a,b}$ 89.4 ± 29.0^{b} 11.8 ± 9.4^{c}						

(or excess). Our present data generally support this; in children within the suggested adequate UIC range of 100-299 μ g/L, the 3rd and 97th percentiles were 3 and 44 μ g/L, respectively, and the prevalence of elevated Tg values was low (1.3% and 2.1%, respectively). Another method of interpreting Tg concentrations in population iodine monitoring is comparison of the median Tg concentration with a cutoff. In 1994, WHO proposed that a median serum Tg concentration <10 µg/L in a population indicated iodine sufficiency (21). However, because there was little evidence to support this Tg cutoff value, the recommendation was not included in later revised WHO guidelines (22). Using clusters with a median UIC in the range of 100–299 μ g/L in our population, we propose that a median of <13μg/L, after CRM-457 standardization, indicates iodine sufficiency in a population of school-aged children. But it is critical that CRM-457 standardization be used when applying this median: using the Tg standards supplied by the manufacturer (PerkinElmer Life Sciences) on DBS, the median Tg concentration of school clusters with median UICs > 100 and < 300 μ g/L was \sim 40% lower. Therefore, it should be emphasized the 13-µg/L cutoff applies specifically to Tg concentrations from assays after CRM-457 standardization.

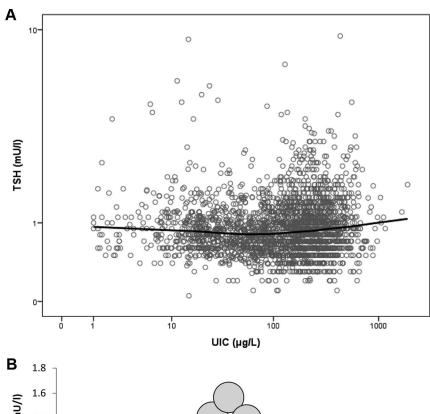
This proposed median Tg cutoff is supported by data from several intervention studies. In a study in a severely deficient area of Morocco (16), before introduction of iodized salt, median DBS-Tg was high (49 µg/L), and more than two-thirds of children had DBS-Tg values greater than 40 μ g/L. After 10 months of iodized salt use, the median had decreased to 8 µg/L and only 3% of children had a value greater than 40 μ g/L. In another study, provision of iodized salt for 12 months in iodine-deficient children reduced median DBS-Tg from a baseline of 25 to 4 μg/L (15). In mildly iodine-deficient New Zealand school-aged children, daily iodine supplementation for 28 weeks raised median UIC from 66 to 145 μ g/L and mean serum Tg decreased from 16 to 9 µg/L (14). In mild and moderately iodine-deficient Danish adults, after the introduction of iodized salt, the median serum Tg significantly decreased from 11 to 9 µg/L in the area with previous mild deficiency and from 15 to 9 μ g/L in the area with previous moderate deficiency; overall, the prevalence of Tg >40 μ g/L fell from 11.3% to 3.7% (13).

Table 3. Prevalence of Thyroid Dysfunction and Percentage of Abnormal Values for Tg, Tg-Ab, and TPO-Ab in 6- to 12-Year-Old Children by WHO Categories of UIC

	UIC, μg/L				
	<50	50-99.9	100-199.9	200-299.9	>300
n	600	358	609	468	477
Subclinical hypothyroidism, % (n)	1.8 (11) ^a	0.3 (1) ^a	0.5 (3) ^a	0.2 (1) ^a	0.6 (3) ^a
Overt hypothyroidism, % (n)	0.7 (4)a	0 (0) ^a	0 (0) ^a	0 (0) ^a	0 (0) ^a
Subclinical hyperthyroidism, % (n)	0.2 (1) ^a	0.3 (1) ^a	0.3 (2) ^a	0 (0) ^a	0.2 (1) ^a
Overt hyperthyroidism, % (n)	0 (0) ^a	0 (0) ^a	0 (0) ^a	0 (0) ^a	0 (0) ^a
Elevated Tg (>40 μ g/L), % (n)	28.8 (173) ^a	5.9 (21) ^b	2.1 (13) ^c	1.3 (6) ^c	8.6 (41) ^b
Elevated Tg Ab (>10 U/mL), % (n)	0 (0) ^a	1.1 (1)a	0.8 (2)a	1.2 (2) ^a	1.0 (2) ^a
Elevated TPO Ab (>12 U/mL), % (n)	0 (0) ^a	0 (0) ^a	0.5 (1) ^a	0.6 (1) ^a	0.5 (1) ^a

Subclinical hypothyroidism as TSH >3.7 mU/L and normal T_4 ; overt hypothyroidism as TSH >3.7 mU/L and T_4 <65 nmol/L; subclinical hyperthyroidism as TSH <0.2 mU/L and normal T_4 ; and overt hyperthyroidism as TSH <0.2 mU/L and T_4 >165 nmol/L.

^{a-c} Means not sharing a common superscript letter are significantly different from each other (χ^2 test followed by z-test; P < .05 was considered significant).



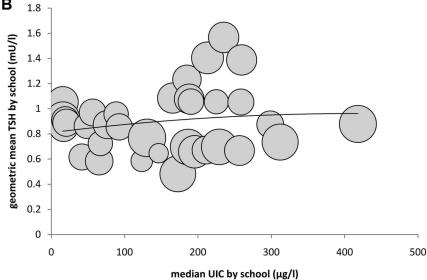


Figure 1. A, Scatterplot (using individual values of 2512 children age 6 to 12 years from 12 countries) of TSH vs UIC with a Loess smoothed line added to show best fit. Data are presented on a log scale for TSH and UIC. B, Bubble plot (clustered by school) of geometric mean TSH vs median UIC with a second-order polynomial trend line. The size of the bubbles reflects the sample size for each school.

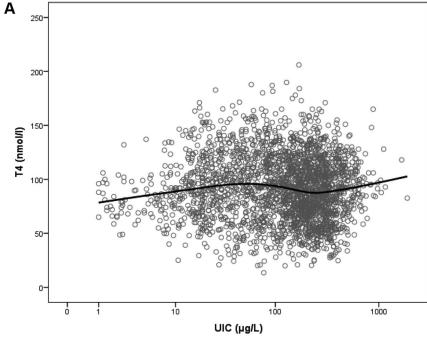
Even with CRM-457 standardization, presumably due to epitope specificity differences that cause interassay biases independent of standardization, there is significant variability between different serum Tg assays that precludes the use of serial serum Tg measurements by different laboratories. This has been demonstrated in clinical practice when following individuals after surgery for differentiated thyroid cancer (20). However, for the purposes of iodine monitoring, ie, distinguishing iodine-deficient from iodine-sufficient populations using a normal refer-

ence interval, assay bias and imprecision goals need not be as stringent as for serial measurements for differentiated thyroid cancer follow-up. Therefore, although use of CRM-457 standardization will not eliminate Tg interassay variability, it may improve the calibration of assays to allow different DBS or serum Tg assays to be used interchangeably to characterize iodine status in a population.

Our data suggest that thyroid Abs are rare in school-aged children and are not increased by either iodine deficiency or excess. Most adult studies report an increase in thyroid autoimmunity at high iodine intake (23-27). Autoimmune thyroiditis may be more common in areas of adequate iodine intake than in areas of iodine deficiency (28-31). In children, the link between iodine intake and thyroid autoimmunity is unclear, but in general, thyroid antibodies are rare in childhood; reported prevalence varies from 0.5%-3% in areas not affected by iodine deficiency disorders to 10%-16% in children and adolescents with goiter (32-37). Previous intervention studies in children with iodized oil (11) and iodized salt (15) found no induction of antithyroid antibodies with higher iodine intakes. A potential limitation to the use of a Tg assay for iodine deficiency disorder monitoring is interference from Tg-Ab (20), but in our sample of children, <1% had elevated anti-Tg-Ab. Thus, screening for Tg-Ab appears unnecessary when using a Tg assay in children to classify population iodine status,

even at excess iodine intake.

In areas of iodine sufficiency, most healthy adults are remarkably tolerant to iodine intake up to 1 mg/d because the thyroid is able to adjust to a wide range of intake to regulate the synthesis and release of thyroid hormones (38). Although large amounts of iodine given for days to months in healthy subjects have shown few adverse effects (39), in adults with past or present thyroid abnormalities, even modest increases in iodine intake in areas of chronic



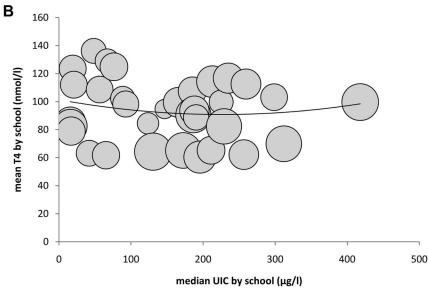


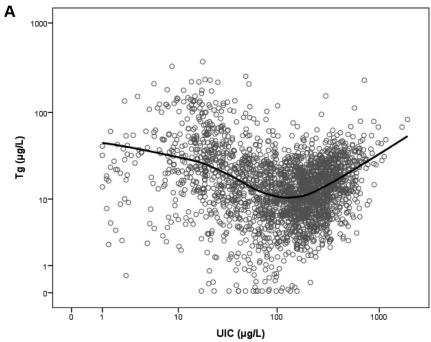
Figure 2. A, Scatterplot (using individual values of 2512 children age 6 to 12 years from 12 countries) of total T_4 vs UIC with a Loess smoothed line added to show best fit. Data are presented on a log scale for UIC. B, Bubble plot (clustered by school) of mean T_4 vs median UIC with a second-order polynomial trend line. The size of the bubbles reflects the sample size for each school.

iodine deficiency can rarely precipitate hyper- or hypothyroidism (2). The fetus and the newborn are particularly vulnerable to iodine excess, and excessive maternal iodine intake can cause neonatal goiter and hypothyroidism (40). In children, excess dietary iodine has been associated with goiter and thyroid dysfunction. In coastal Japan (41), consumption of iodine-rich seaweed with intake of >20 mg iodine per day was associated with a prevalence of visible goiter in children of 3%–9%, but no cases of clinical hypoor hyperthyroidism were reported. In Chinese children consuming iodine-rich drinking water (462 µg/L) with a

mean UIC of 1235 µg/g creatinine, mean serum TSH was elevated at 7.8 mU/L, and the goiter rate was >60%(42). In other reports from China, drinking water with iodine concentrations >300 μ g/L resulted in UIC >900 μ g/L and a goiter rate >10% (43). In a large international study of 6- to 12-year-old children, chronic iodine intakes \geq 500 µg/d were associated with an increase in thyroid size by ultrasonography (44). These past studies suggest goiter begins to appear in children when iodine intake increases above 400-500 μg/d. In our study, the higher frequency of children with an elevated Tg above the 300-µg/L UIC threshold suggests the onset of thyroid hyperstimulation.

Establishing the ideal range of values for urinary iodine for monitoring is difficult. A USI program should be able to meet the increased needs of pregnant and lactating women without supplying too much iodine to other population groups (3). The median UIC that indicates sufficient iodine intake in pregnant women is $150-250 \mu g/L$, but at the same time, it is recommended that the median UIC for school-aged children be kept in the range of $100-199 \mu g/L$. Because daily urine volumes are higher in pregnant women, this leaves a relatively narrow range for iodine intake that will both meet the increased needs for pregnant/lactating women and not be excessive for school-aged children. In our data, there was no change in the prevalence of elevated Tg (or antithyroid Abs) comparing children across the WHO ranges of

adequate (UIC range of $100-199~\mu g/L$) and more than adequate iodine intake (UIC range of $200-299~\mu g/L$). These findings indicate iodine intakes resulting in UICs in the current WHO category of more than adequate intake $(200-299~\mu g/L)$ do not cause thyroid dysfunction in children. Thus, it may be prudent to widen the acceptable range of median UIC for children and consider adoption of a single category of sufficient iodine intake in the range of $100-299~\mu g/L$ for children in revised program monitoring guidelines. There was, however, a significant in-



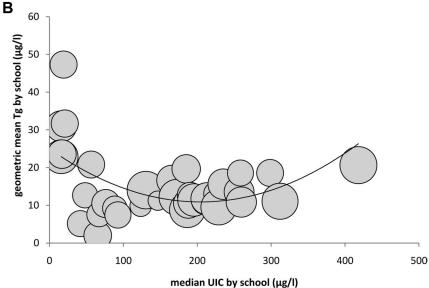


Figure 3. A, Scatterplot (using individual values of 2512 children age 6 to 12 years from 12 countries) of DBS Tg vs UIC with a Loess smoothed line added to show best fit. Data are presented on a log scale for Tg and UIC. B, Bubble plot (clustered by school) of geometric mean Tg vs median UIC with a second-order polynomial trend line. The size of the bubbles reflects the sample size for each school.

crease in the prevalence of elevated Tg in the UIC ranges of 50-99 and $<50 \,\mu\text{g/L}$. Thus, the data support the use of a median UIC $<100 \,\mu\text{g/L}$ to define iodine deficiency in populations of school-age children.

The strengths of this study include 1) the use of identical assays for all thyroid function tests and the same method for all urinary iodine measurements with external quality control, 2) its large international and multiethnic sample, and 3) the fact that the high iodine intake in the children was due to dietary sources of iodine including iodized salt. These make the results generalizable in the global context of monitoring of salt iodization programs. However, in-

terpretation of epidemiological studies linking iodine intake and thyroid function is challenging, for several reasons. One should consider not only the present iodine intake level but also the history of iodine intake of the population. In our sample, we collected data from regions where children had been chronically exposed to a constant level of iodine intake over many years. Unmeasured environmental factors (eg, goitrogens and micronutrient status) (45) as well as differing genetic background could have modulated the relationship between iodine intake and thyroid function. But overall, our data indicate Tg may be a useful biological indicator for monitoring thyroid function in children after introduction of iodized salt when used together with UIC to assess recent iodine intake.

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References

- Andersson M, Karumbunathan V, Zimmermann MB. Global iodine status in 2011 and trends over the past decade. J Nutr. 2012;142(4): 744-750.
- 2. Zimmermann MB. Iodine deficiency. Endocr Rev. 2009;30(4):376–408.
- WHO/UNICEF/ICCIDD. Assessment of Iodine Deficiency Disorders and Monitoring Their Elimination. 3rd ed. Geneva, Switzerland: World Health Organization; 2007.
- Torrens JI, Burch HB. Serum thyroglobulin measurement: utility in clinical practice. *Endocrinol Metab Clin North Am.* 2001;30(2): 429–467.
- 5. de Vijlder JJM, Ris-Stalpers C, Vulsma T. On the origin of circulating thyroglobulin. *Eur J Endocrinol*. 1999;140(1):7–8.
- Targovnik HM. Thyroglobulin: structure, function, and biosynthesis. In: Braverman LE, Cooper DS, eds. Werner and Ingbar's The Thyroid: A Fundamental and Clinical Text. Philadelphia, PA: Lippincott Williams & Wilkins; 2013:74–92.
- Spencer CA, Wang CC. Thyroglobulin measurement: techniques, clinical benefits, and pitfalls. *Endocrinol Metab Clin North Am*. 1995;24(4):841.
- Knudsen N, Bulow I, Jorgensen T, Perrild H, Ovesen L, Laurberg P. Serum Tg: a sensitive marker of thyroid abnormalities and iodine deficiency in epidemiological studies. *J Clin Endocrinol Metab*. 2001;86(8):3599–3603.
- van den Briel T, West CE, Hautvast JG, Vulsma T, de Vijlder JJ, Ategbo EA. Serum thyroglobulin and urinary iodine concentration are the most appropriate indicators of iodine status and thyroid function under conditions of increasing iodine supply in schoolchildren in Benin. J Nutr. 2001;131(10):2701–2706.
- Missler U, Gutekunst R, Wood WG. Thyroglobulin is a more sensitive indicator of iodine deficiency than thyrotropin: development and evaluation of dry blood spot assays for thyrotropin and thyroglobulin in iodine-deficient geographical areas. *Eur J Clin Chem Clin Biochem.* 1994;32(3):137–143.
- Benmiloud M, Chaouki ML, Gutekunst R, Teichert HM, Wood WG, Dunn JT. Oral iodized oil for correcting iodine deficiency: optimal dosing and outcome indicator selection. *J Clin Endocrinol Metab*. 1994;79(1):20–24.
- Todd CH, Dunn JT. Intermittent oral administration of potassium iodide solution for the correction of iodine deficiency. *Am J Clin Nutr.* 1998;67(6):1279–1283.
- 13. Vejbjerg P, Knudsen N, Perrild H, et al. Thyroglobulin as a marker of iodine nutrition status in the general population. *Eur J Endocrinol*. 2009;161(3):475–481.
- Gordon RC, Rose MC, Skeaff SA, Gray AR, Morgan KM, Ruffman T. Iodine supplementation improves cognition in mildly iodine-deficient children. *Am J Clin Nutr.* 2009;90(5):1264–1271.
- 15. Zimmermann MB, Moretti D, Chaouki N, Torresani T. Development of a dried whole-blood spot thyroglobulin assay and its evaluation as an indicator of thyroid status in goitrous children receiving iodized salt. *Am J Clin Nutr.* 2003;77(6):1453–1458.
- 16. Zimmermann MB, de Benoist B, Corigliano S, et al. Assessment of iodine status using dried blood spot thyroglobulin: development of reference material and establishment of an international reference range in iodine-sufficient children. J Clin Endocrinol Metab. 2006; 91(12):4881–4887.

- 17. Armitage P, Berry G, Matthews JN. Statistical Methods in Medical Research. 4th ed. Oxford, UK: Blackwell Science; 2002.
- 18. World Health Organization. *Physical Status: The Use and Interpretation of Anthropometry*. Report of a WHO Expert Committee. Geneva, Switzerland: World Health Organization; 1995. Technical Report Series No. 854.
- 19. Pino S, Fang SL, Braverman LE. Ammonium persulfate: a safe alternative oxidizing reagent for measuring urinary iodine. *Clin Chem.* 1996;42(2):239–243.
- Demers LM, Spencer CA. Laboratory support for the diagnosis and monitoring of thyroid disease: thyroglobulin (Tg) measurement. Laboratory Medicine Practice Guidelines: The National Academy of Clinical Biochemistry. 2002:57–67.
- 21. WHO/ICCIDD/UNICEF. Indicators for Assessing Iodine Deficiency Disorders and Their Control Through Salt Iodization. Geneva, Switzerland: World Health Organization; 1994.
- 22. WHO/UNICEF/ICCIDD. Assessment of Iodine Deficiency Disorders and Monitoring Their Elimination: A Guide for Programme Managers. 2nd ed. Geneva, Switzerland: World Health Organization; 2001.
- 23. Boukis MA, Koutras DA, Souvatzoglou A, Evangelopoulou A, Vrontakis M, Moulopoulos SD. Thyroid-hormone and immunological studies in endemic goiter. *J Clin Endocrinol Metab.* 1983; 57(4):859–862.
- 24. Kahaly GJ, Dienes HP, Beyer J, Hommel G. Iodide induces thyroid autoimmunity in patients with endemic goitre: a randomised, double-blind, placebo-controlled trial. *Eur J Endocrinol*. 1998;139(3): 290–297.
- Koutras DA, Piperingos GD, Pallas D, et al. Clinical, laboratory and immunologic effects of the treatment of endemic goiter with T₄, T₃ and KI. *Thyroidology*. 1990;2(2):81–88.
- 26. Pedersen IB, Knudsen N, Carle A, Vejbjerg P, Jorgensen T, Perrild H, Ovesen L, Rasmussen LB, Laurberg P. A cautious iodization programme bringing iodine intake to a low recommended level is associated with an increase in the prevalence of thyroid autoantibodies in the population. *Clin Endocrinol.* 2011;75(1):120–126.
- 27. Laurberg P, Cerqueira C, Ovesen L, et al. Iodine intake as a determinant of thyroid disorders in populations. *Best Pract Res Clin Endocrinol Metab*. 2010;24(1):13–27.
- 28. Dayan CM, Daniels GH. Chronic autoimmune thyroiditis. *N Engl J Med*. 1996;335(2):99–107.
- 29. Weetman AP, Mcgregor AM. Autoimmune thyroid disease: further developments in our understanding. *Endocr Rev.* 1994;15(6):788–830
- Konno N, Makita H, Yuri K, Iizuka N, Kawasaki K. Association between dietary iodine intake and prevalence of subclinical hypothyroidism in the coastal regions of Japan. *J Clin Endocrinol Metab*. 1994;78(2):393–397.
- Laurberg P, Pedersen KM, Hreidarsson A, Sigfusson N, Iversen E, Knudsen PR. Iodine intake and the pattern of thyroid disorders: a comparative epidemiological study of thyroid abnormalities in the elderly in Iceland and in Jutland, *Denmark. J Clin Endocrinol* Metab. 1998;83(3):765–769.
- 32. Tsatsoulis A, Johnson EO, Andricula M, Kalogera C, Svarna E, Spyroy P, Seferiadis K, Tsolas O. Thyroid autoimmunity is associated with higher urinary iodine concentrations in an iodine-deficient area of Northwestern Greece. *Thyroid*. 1999;9(3):279–283.
- 33. Marwaha RK, Tandon N, Karak AK, Gupta N, Verma K, Kochupillai N. Hashimoto's thyroiditis: Countrywide screening of goitrous healthy young girls in postiodization phase in India. *J Clin Endocrinol Metab.* 2000;85(10):3798–3802.
- 34. Jaksic J, Dumic M, Filipovic B, Ille J, Cvijetic M, Gjuric G. Thyroid diseases in a school population with thyromegaly. *Arch Dis Child*. 1994;70(2):103–106.
- 35. Jaruratanasirikul S, Sopanapikul S, Mo-Suwan L, et al. Goiter in Thai schoolchildren: study in Hat Yai, southern Thailand. *J Med Assoc Thai*. 1995;78(9):449–454.

- 36. Wong GW, Lam CW, Kwok MY, et al. Childhood goitre and urinary iodine excretion in Hong Kong. *Eur J Pediatr*. 1998;157(1):8–12.
- 37. Aghini-Lombardi F, Antonangeli L, Martino E, et al. The spectrum of thyroid disorders in an iodine-deficient community: the Pescopagano survey. *J Clin Endocrinol Metab*. 1999;84(2):561–566.
- 38. Chow CC, Phillips DI, Lazarus JH, Parkes AB. Effect of low-dose iodide supplementation on thyroid function in potentially susceptible subjects: are dietary iodide levels in Britain acceptable. *Clin Endocrinol (Oxf)*. 1991;34(5):413–416.
- Gardner DF, Centor RM, Utiger RD. Effects of low-dose oral iodide supplementation on thyroid function in normal men. *Clin Endocri*nol (Oxf). 1988;28(3):283–288.
- Emder PJ, Jack MM. Iodine-induced neonatal hypothyroidism secondary to maternal seaweed consumption: a common practice in

- some Asian cultures to promote breast milk supply. *J Paediatr Child Health*. 2011;47(10):750–752.
- 41. Suzuki H, Higuchi T, Sawa K, Ohtaki S, Horiuchi Y. "Endemic coast goitre" in Hokkaido, Japan. *Acta Endocrinol (Copenh)*. 1965; 50(2):161–176.
- 42. Li M, Liu DR, Qu CY, et al. Endemic goitre in central China caused by excessive iodine intake. *Lancet*. 1987;2(8553):257–259.
- 43. Zhao J, Chen Z, Maberly G. Iodine-rich drinking water of natural origin in China. *Lancet*. 1998;352(9145):2024.
- 44. Zimmermann MB, Ito Y, Hess SY, Fujieda K, Molinari L. High thyroid volume in children with excess dietary iodine intakes. *Am J Clin Nutr*. 2005;81(4):840–844.
- 45. **Zimmermann MB.** The influence of iron status on iodine utilization and thyroid function. *Annu Rev Nutr.* 2006;26:367–389.