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Persistence of coronaviruses on inanimate surfaces and its inactivation with biocidal agents

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## Summary

Currently, the emergence of a novel human coronavirus, temporary named 2019-nCoV, has become a global health concern causing severe respiratory tract infections in humans. Human-to-human transmissions have been described with incubation times between 2-10 days, facilitating its spread via droplets, contaminated hands or surfaces. We therefore reviewed the literature on all available information about the persistence of human and veterinary coronaviruses on inanimate surfaces as well as inactivation strategies with biocidal agents used for chemical disinfection, e.g. in healthcare facilities. The analysis of 22 studies reveals that human coronaviruses such as Severe Acute Respiratory Syndrome (SARS) coronavirus, Middle East Respiratory Syndrome (MERS) coronavirus or endemic human coronaviruses (HCoV) can persist on inanimate surfaces like metal, glass or plastic for up to 9 days, but can be efficiently inactivated by surface disinfection procedures with 62-71% ethanol, 0.5% hydrogen peroxide or 0.1% sodium hypochlorite within 1 minute. Other biocidal agents such as 0.05-0.2% benzalkonium chloride or 0.02% chlorhexidine digluconate are less effective. As no specific therapies are available for 2019-nCoV, early containment and prevention of further spread will be crucial to stop the ongoing outbreak and to control this novel infectious thread.

**Keywords:** coronavirus; persistence; inanimate surfaces; chemical inactivation; biocidal agents; disinfection

## Introduction

A novel coronavirus (2019-nCoV) has recently emerged from China with a total of 6065 laboratory-confirmed cases of pneumonia (as of January 30, 2020) [1]. Together with Severe Acute Respiratory Syndrome (SARS) coronavirus and Middle East Respiratory Syndrome (MERS) coronavirus [2], this is the third highly pathogenic human coronavirus that has emerged in the last two decades. Person-to-person transmission has been described both in hospital and family settings [3]. It is therefore of utmost importance to prevent any further spread in the public and healthcare settings. Transmission of coronaviruses from contaminated dry surfaces has been postulated including self-inoculation of mucous membranes of the nose, eyes or mouth [4, 5], emphasizing the importance of a detailed understanding of coronavirus persistence on inanimate surfaces [6]. Various types of biocidal agents such as hydrogen peroxide, alcohols, sodium hypochlorite or benzalkonium chloride are used worldwide for disinfection, mainly in healthcare settings [7]. The aim of the review was therefore to summarize all available data on the persistence of all coronaviruses including emerging SARS-CoV and MERS-CoV as well as veterinary coronaviruses such as transmissible gastroenteritis virus (TGEV), mouse hepatitis virus (MHV) and canine coronavirus (CCV) on different types of inanimate surfaces and on the efficacy of commonly used biocidal agents used in surface disinfectants against coronaviruses.

## Method

A Medline search has been done on January 28, 2020. The following terms were used, always in combination with "coronavirus", "TGEV", "MHV" and "CCV": survival surface (88 / 10 / 25 / 0 hits), persistence surface (47 / 1 / 32 / 0 hits), persistence hand (8 / 0 / 3 / 0 hits), survival hand (22 / 0 / 3 / 1 hits), survival skin (8 / 0 / 0 / 1 hits), persistence skin (1 / 0 / 0 / 1 hit), virucidal (23 / 3 / 3 / 1 hits), chemical inactivation (33 / 0 / 6 / 1), suspension test (18 / 0 / 0 / 0 hits) and carrier test (17 / 4 / 0 / 0). Publications were included and results were extracted given they provided original data on coronaviruses on persistence (surfaces, materials) and inactivation by biocidal agents used for disinfection (suspension tests, carrier tests, fumigation studies). Data with commercial products based on various different types of biocidal agents were excluded. Reviews were not included, but screened for any information within the scope of this review.

## Results

### *Persistence of coronavirus on inanimate surfaces*

Most data were described with the endemic human coronavirus strain (HCoV-) 229E. On different types of materials it can remain infectious for from 2 hours up to 9 days. A higher temperature such as 30°C or 40°C reduced the duration of persistence of highly pathogenic MERS-CoV, TGEV and MHV. However, at 4°C persistence of TGEV and MHV can be increased to  $\geq 28$  days. Few comparative data obtained with SARS-CoV indicate that persistence was longer with higher inocula (Table I). In addition it was shown at room temperature that HCoV-229E persists better at 50% compared to 30% relative humidity [8].

### *Inactivation of coronaviruses by biocidal agents in suspension tests*

Ethanol (78-95%), 2-propanol (70-100%), the combination of 45% 2-propanol with 30% 1-propanol, glutardialdehyde (0.5-2.5%), formaldehyde (0.7-1%) and povidone iodine (0.23-7.5%) readily inactivated coronavirus infectivity by approximately 4 log<sub>10</sub> or more. (Table 2). Sodium hypochlorite required a minimal concentration of at least 0.21% to be effective. Hydrogen peroxide was effective with a concentration of 0.5% and an incubation time of 1 min. Data obtained with benzalkonium chloride at reasonable contact times were conflicting. Within 10 min a concentration of 0.2%

revealed no efficacy against coronavirus whereas a concentration of 0.05% was quite effective. 0.02% chlorhexidine digluconate was basically ineffective (Table II).

#### *Inactivation of coronaviruses by biocidal agents in carrier tests*

Ethanol at concentrations between 62% and 71% reduced coronavirus infectivity within 1 min exposure time by 3.0–4.0  $\log_{10}$ . Concentrations of 0.1–0.5% sodium hypochlorite and 2% glutaraldehyde were also quite effective with  $> 3.0 \log_{10}$  reduction in viral titre. In contrast, 0.04% benzalkonium chloride, 0.06% sodium hypochlorite and 0.55% ortho-phthalaldehyde were less effective (Table III).

#### **Discussion**

Human coronaviruses can remain infectious on inanimate surfaces at room temperature for up to 9 days. At a temperature of 30°C or more the duration of persistence is shorter. Veterinary coronaviruses have been shown to persist even longer for 28 d. Contamination of frequent touch surfaces in healthcare settings are therefore a potential source of viral transmission. Data on the transmissibility of coronaviruses from contaminated surfaces to hands were not found. However, it could be shown with influenza A virus that a contact of 5 s can transfer 31.6% of the viral load to the hands [9]. The transfer efficiency was lower (1.5%) with parainfluenza virus 3 and a 5 s contact between the surface and the hands [10]. In an observational study, it was described that students touch their face with their own hands on average 23 times per h, with contact mostly to the skin (56%), followed by mouth (36%), nose (31%) and eyes (31%) [11]. Although the viral load of coronaviruses on inanimate surfaces is not known during an outbreak situation it seems plausible to reduce the viral load on surfaces by disinfection, especially of frequently touched surfaces in the immediate patient surrounding where the highest viral load can be expected. The WHO recommends “to ensure that environmental cleaning and disinfection procedures are followed consistently and correctly. Thoroughly cleaning environmental surfaces with water and detergent and applying commonly used hospital-level disinfectants (such as sodium hypochlorite) are effective and sufficient procedures.” [12] The typical use of bleach is at a dilution of 1:100 of 5% sodium hypochlorite resulting in a final concentration of 0.05% [13]. Our summarized data with coronaviruses suggest that a concentration of 0.1% is effective in 1 min (Table 3). That is why it seems appropriate to recommend a dilution 1:50 of standard bleach in the coronavirus setting. For the disinfection of small surfaces ethanol (62–71%; carrier tests) revealed a similar efficacy against coronavirus. A concentration of 70% ethanol is also recommended by the WHO for disinfecting small surfaces [13].

No data were found to describe the frequency of hands becoming contaminated with coronavirus, or the viral load on hands either, after patient contact or after touching contaminated surfaces. The WHO recommends to preferably apply alcohol-based hand rubs for the decontamination of hands, e.g. after removing gloves. Two WHO recommended formulations (based on 80% ethanol or 75% 2-propanol) have been evaluated in suspension tests against SARS-CoV and MERS-CoV, and both were described to be very effective [14]. No in vitro data were found on the efficacy of hand washing against coronavirus contaminations on hands. In Taiwan, however, it was described that installing hand wash stations in the emergency department was the only infection control measure which was significantly associated with the protection from healthcare workers from acquiring the SARS-CoV, indicating that hand hygiene can have a protective effect [15]. Compliance with hand hygiene can be significantly higher in an outbreak situation but is likely to remain an obstacle especially among physicians [16–18]. Transmission in healthcare settings can be successfully prevented when appropriate measures are consistently performed [19, 20].

#### **Conclusions**

Human coronaviruses can remain infectious on inanimate surfaces for up to 9 days. Surface disinfection with 0.1% sodium hypochlorite or 62-71% ethanol significantly reduces coronavirus infectivity on surfaces within 1 min exposure time. We expect a similar effect against the 2019-nCoV.

#### Conflict of interest

None.

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**Table I.** Persistence of coronaviruses on different types of inanimate surfaces.

Type of surface	Virus	Strain / isolate	Inoculum (viral titer)	Temperature	Persistence	Reference
Steel	MERS-CoV	Isolate HCoV-EMC/2012	$10^5$	20°C	48 h	[21]
				30°C	8 – 24 h	
	TGEV	Unknown	$10^6$	4°C	$\geq 28$ d	[22]
				20°C	3 – 28 d	
	MHV	Unknown	$10^6$	40°C	4 – 96 h	[22]
				4°C	$\geq 28$ d	
Aluminium	HCoV	Strain 229E	$10^3$	20°C	4 – 28 d	[23]
				40°C	4 – 96 h	
Metal	SARS-CoV	Strain P9	$10^5$	21°C	5 d	[23]
Wood	SARS-CoV	Strain P9	$10^5$	RT	2 – 8 h	[24]
					5 d	[25]
Paper	SARS-CoV	Strain P9	$10^5$	RT	4 d	[25]
					4 – 5 d	[25]
					24 h	[26]
Glass	SARS-CoV	Strain P9	$10^5$	RT	3 h	[26]
					$10^4$	< 5 min
Plastic	SARS-CoV	Strain P9	$10^5$	RT	4 d	[25]
					HCoV	Strain 229E
Plastic	SARS-CoV	Strain HKU39849	$10^5$	22°-25°C	$\leq 5$ d	[27]
					MERS-CoV	Isolate HCoV-EMC/2012
	SARS-CoV	Strain P9	$10^5$	30°C	8 – 24 h	[21]
	SARS-CoV	Strain FFM1	$10^7$	RT	4 d	[25]
PVC	HCoV	Strain 229E	$10^7$	RT	6 – 9 d	[28]
					2 – 6 d	[28]
Silicon rubber	HCoV	Strain 229E	$10^3$	RT	5 d	[23]
Surgical glove (latex)	HCoV	Strain 229E	$10^3$	21°C	5 d	[23]
Disposable gown	SARS-CoV	Strain GUV6109	$10^3$	21°C	$\leq 8$ h	[24]
					$10^6$	2 d
			$10^5$	RT	24 h	[26]

			$10^4$		1 h	
Ceramic	HCoV	Strain 229E	$10^3$	21°C	5 d	[23]
Teflon	HCoV	Strain 229E	$10^3$	21°C	5 d	[23]

MERS = Middle East Respiratory Syndrome; HCoV = human coronavirus; TGEV = transmissible gastroenteritis virus; MHV = mouse hepatitis virus; SARS = Severe Acute Respiratory Syndrome; RT = room temperature.

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**Table II.** Inactivation of coronaviruses by different types of biocidal agents in suspension tests.

Biocidal agent	Concentration	Virus	Strain / isolate	Exposure time	Reduction of viral infectivity ( $\log_{10}$ )	Reference
Ethanol	95%	SARS-CoV	Isolate FFM-1	30 s	$\geq 5.5$	[29]
	85%	SARS-CoV	Isolate FFM-1	30 s	$\geq 5.5$	[29]
	80%	SARS-CoV	Isolate FFM-1	30 s	$\geq 4.3$	[29]
	80%	MERS-CoV	Strain EMC	30 s	$> 4.0$	[14]
	78%	SARS-CoV	Isolate FFM-1	30 s	$\geq 5.0$	[28]
	70%	MHV	Strains MHV-2 and MHV-N	10 min	$> 3.9$	[30]
	70%	CCV	Strain I-71	10 min	$> 3.3$	[30]
2-Propanol	100%	SARS-CoV	Isolate FFM-1	30 s	$\geq 3.3$	[28]
	75%	SARS-CoV	Isolate FFM-1	30 s	$\geq 4.0$	[14]
	75%	MERS-CoV	Strain EMC	30 s	$\geq 4.0$	[14]
	70%	SARS-CoV	Isolate FFM-1	30 s	$\geq 3.3$	[28]
	50%	MHV	Strains MHV-2 and MHV-N	10 min	$> 3.7$	[30]
	50%	CCV	Strain I-71	10 min	$> 3.7$	[30]
2-Propanol and 1-propanol	45% and 30%	SARS-CoV	Isolate FFM-1	30 s	$\geq 4.3$	[29]
		SARS-CoV	Isolate FFM-1	30 s	$\geq 2.8$	[28]
Benzalkonium chloride	0.2%	HCoV	ATCC VR-759 (strain OC43)	10 min	0.0	[31]
	0.05%	MHV	Strains MHV-2 and MHV-N	10 min	$> 3.7$	[30]
	0.05%	CCV	Strain I-71	10 min	$> 3.7$	[30]
	0.00175%	CCV	Strain S378	3 d	3.0	[32]
Didecyldimethyl ammonium chloride	0.0025%	CCV	Strain S378	3 d	$> 4.0$	[32]
Chlorhexidine digluconate	0.02%	MHV	Strains MHV-2 and MHV-N	10 min	0.7 – 0.8	[30]
	0.02%	CCV	Strain I-71	10 min	0.3	[30]
Sodium hypochlorite	0.21%	MHV	Strain MHV-1	30 s	$\geq 4.0$	[33]
	0.01%	MHV	Strains MHV-2 and MHV-N	10 min	2.3 – 2.8	[30]
	0.01%	CCV	Strain I-71	10 min	1.1	[30]
	0.001%	MHV	Strains MHV-2 and MHV-N	10 min	0.3 – 0.6	[30]
	0.001%	CCV	Strain I-71	10 min	0.9	[30]
Hydrogen peroxide	0.5%	HCoV	Strain 229E	1 min	$> 4.0$	[34]
Formaldehyde	1%	SARS-CoV	Isolate FFM-1	2 min	$> 3.0$	[28]

	0.7%	SARS-CoV	Isolate FFM-1	2 min	> 3.0	[28]
	0.7%	MHV		10 min	> 3.5	[30]
	0.7%	CCV	Strain I-71	10 min	> 3.7	[30]
	0.009%	CCV		24 h	> 4.0	[35]
Glutaraldehyde	2.5%	SARS-CoV	Hanoi strain	5 min	> 4.0	[36]
	0.5%	SARS-CoV	Isolate FFM-1	2 min	> 4.0	[28]
Povidone iodine	7.5%	MERS-CoV	Isolate HCoV-EMC/2012	15 s	4.6	[37]
	4%	MERS-CoV	Isolate HCoV-EMC/2012	15 s	5.0	[37]
	1%	SARS-CoV	Hanoi strain	1 min	> 4.0	[36]
	1%	MERS-CoV	Isolate HCoV-EMC/2012	15 s	4.3	[37]
	0.47%	SARS-CoV	Hanoi strain	1 min	3.8	[36]
	0.25%	SARS-CoV	Hanoi strain	1 min	> 4.0	[36]
	0.23%	SARS-CoV	Hanoi strain	1 min	> 4.0	[36]
	0.23%	SARS-CoV	Isolate FFM-1	15 s	≥ 4.4	[38]
	0.23%	MERS-CoV	Isolate HCoV-EMC/2012	15 s	≥ 4.4	[38]

SARS = Severe Acute Respiratory Syndrome; MERS = Middle East Respiratory Syndrome; MHV = mouse hepatitis virus; CCV = canine coronavirus; HCoV = human coronavirus.

**Table III.** Inactivation of coronaviruses by different types of biocidal agents in carrier tests.

Biocidal agent	Concentration	Virus	Strain / isolate	Volume / material	Organic load	Exposure time	Reduction of viral infectivity ( $\log_{10}$ )	Reference
Ethanol	71%	TGEV	Unknown	50 $\mu$ l / stainless steel	None	1 min	3.5	[39]
	71%	MHV	Unknown	50 $\mu$ l / stainless steel	None	1 min	2.0	[39]
	70%	TGEV	Unknown	50 $\mu$ l / stainless steel	None	1 min	3.2	[39]
	70%	MHV	Unknown	50 $\mu$ l / stainless steel	None	1 min	3.9	[39]
	70%	HCoV	Strain 229E	20 $\mu$ l / stainless steel	5% serum	1 min	> 3.0	[40]
	62%	TGEV	Unknown	50 $\mu$ l / stainless steel	None	1 min	4.0	[39]
	62%	MHV	Unknown	50 $\mu$ l / stainless steel	None	1 min	2.7	[39]
Benzalkoniumchloride	0.04%	HCoV	Strain 229E	20 $\mu$ l / stainless steel	5% serum	1 min	< 3.0	[40]
Sodium hypochlorite	0.5%	HCoV	Strain 229E	20 $\mu$ l / stainless steel	5% serum	1 min	> 3.0	[40]
	0.1%	HCoV	Strain 229E	20 $\mu$ l / stainless steel	5% serum	1 min	> 3.0	[40]
	0.06%	TGEV	Unknown	50 $\mu$ l / stainless steel	None	1 min	0.4	[39]
	0.06%	MHV	Unknown	50 $\mu$ l / stainless steel	None	1 min	0.6	[39]
	0.01%	HCoV	Strain 229E	20 $\mu$ l / stainless steel	5% serum	1 min	< 3.0	[40]
Glutardialdehyde	2%	HCoV	Strain 229E	20 $\mu$ l / stainless steel	5% serum	1 min	> 3.0	[40]
Ortho-phtalaldehyde	0.55%	TGEV	Unknown	50 $\mu$ l / stainless steel	None	1 min	2.3	[39]
	0.55%	MHV	Unknown	50 $\mu$ l / stainless steel	None	1 min	1.7	[39]
Hydrogen peroxide	Vapor of unknown concentration	TGEV	Purdue strain type 1	20 $\mu$ l / stainless steel	None	2 – 3 h	4.9 – 5.3*	[41]

TGEV = transmissible gastroenteritis virus; MHV = mouse hepatitis virus; HCoV = human coronavirus; \*depending on the volume of injected hydrogen peroxide